## 2-Oxindole Acts as a Synthon of 2-Aminobenzoyl Anion in the K<sub>2</sub>CO<sub>3</sub>-Catalyzed Reaction with Enones: Preparation of 1,4-Diketones Bearing an Amino Group and Their Further Transformations

Chun-Bao Miao,\* Yu-Mei Zeng, Tong Shi, Rui Liu, Peng-Fei Wei, Xiao-Qiang Sun, and Hai-Tao Yang

Jiangsu Key Laboratory of Advanced Catalytic Materials and Technology, Advanced Catalysis and Green Manufacturing Collaborative Innovation Center, School of Petrochemical Engineering, Changzhou University, Changzhou 213164, China

Supporting Information

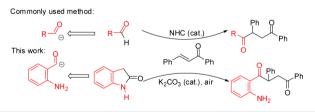
**ABSTRACT:** A convenient approach for the synthesis of 1,4diketones bearing an amino group has been developed through the  $K_2CO_3$ -catalyzed reaction of 2-oxindoles with enones with the assistance of atmospheric  $O_2$  via sequential Michael addition—oxidation—ring-cleavage process. The further intramolecular reaction leads to the formation of benzoazepinone, quinoline, and 3-oxindole derivatives.

# $\begin{array}{c} TSOHH_{2}O\\ (0.1 \text{ equiv})\\ \hline (DMSO, rt, air \\ \hline Michael addition-Oxidation-Ring cleavage \end{array} \xrightarrow{Ph} \\ \hline Michael addition-Oxidation-Ring cleavage \end{array} \xrightarrow{Ph} \\ \hline TSOHH_{2}O\\ (0.1 \text{ equiv})\\ \hline (0.1 \text{ e$

#### INTRODUCTION

1,4-Diketones are useful precursors to a variety of synthetically valuable compounds such as pyrroles, furans, thiophenes,<sup>1</sup> pyridazines,<sup>2</sup> and cyclopentenones.<sup>3</sup> A number of reactions have been developed to construct this important structural pattern.<sup>4</sup> Among them, the conjugate addition of aldehydes onto  $\alpha,\beta$ -unsaturated ketones catalyzed by *N*-heterocyclic carbenes (NHCs), known as the Stetter reaction, was one of the most important methods to access 1,4-diketone skeletons (Scheme 1).<sup>5</sup> An acyl anion equivalent was generated through

Scheme 1. 2-Oxindole as an Equivalent of *o*-Aminobenzoyl Anion



umpolung of aldehydes catalyzed by NHCs, which was often generated in situ from a five-membered nitrogen-containing heterocyclic core like thiazolium, triazolium, imidazolium, and imidazolinium salts in the presence of a base. Recently, the Gravel group developed a novel bis(amino)cyclopropenylidene catalyst as a candidate of NHCs, which even showed high efficiency toward those well-known unreactive substrates in the Stetter reaction catalyzed by NHCs.<sup>6</sup> Notably, a drawback of the Stetter reaction was that most of the aldehyde moieties could not contain amino or hydroxyl groups, which reduced the opportunity for further functionalization of the generated products. 3-Monosubstituted 2-oxindole is a good nucleophile for Michael addition<sup>7</sup> and Michael addition initiated cascade reactions.<sup>8</sup> In terms of 3-unsubstituted oxindole, it is liable to take place condensation reaction with ketonic compounds<sup>9</sup> or double Michael addition reaction with bisenones<sup>10</sup> to form 3-alkylideneoxindoles or spirooxindoles, respectively. The Michael addition reaction of 3-unsubstituted 2-oxindole with enone to give 3-monosubstituted 2-oxindole has only been reported in a few literature refrences in the presence of sodium ethoxide, piperidine, or proline.<sup>11</sup> Here, we report a base-catalyzed reaction of 2-oxindoles with enones for the preparation of 1,4-diketones bearing an amino group through a cascade Michael addition—oxidation—ring-cleavage process. The 2-oxindole was used as an equivalent of *o*-aminobenzoyl anion for the first time (Scheme 1).

#### RESULTS AND DISCUSSION

Initially, the  $K_2CO_3$ -catalyzed addition reaction of oxindole 1a with chalcone 2a was carried out in ethanol at room temperature (Table 1). The Michael adducts 3a and 3b were obtained as the main products 1 h later.<sup>12</sup> With the extension of reaction time, two new products 4aa and 5 were generated, and the amounts of 3a and 3b decreased gradually. The 4aa and 5 could be isolated in 30% and 6% yield 48 h later, respectively. Undoubtedly, the ethoxyl in the carbamate group of 5 was derived from the ethanol solvent. Changing the solvent from ethanol to methanol gave similar results. The product 6 was produced instead of 5, and the ratio of 4/6 is about 2.5 times of the ratio of 4/5.

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#### Table 1. Primary Results

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				yield <sup>a</sup> (%)			
entry	solvent	time (h)	3a + 3b	4aa	5 or 6		
1	EtOH	1	71	trace	0		
2	EtOH	6	78	5	trace		
3	EtOH	24	61	18	3		
4	EtOH	48	49	30	6		
5	MeOH	1	80	6	trace		
6	MeOH	6	69	12	4		
7		24	<b>C</b> 4	19	10		
/	MeOH	24	54	19	10		

The product **4aa** looked like a Michael adduct of *o*aminobenzoyl anion with **2a**. To the best of our knowledge, the umpolung of 2-aminobenzaldehyde to the *o*-aminobenzoyl anion in the Stetter reaction has never been realized. On the other hand, an oxidative process was involved with  $O_2$  as the oxidant, and only a catalytic amount of  $K_2CO_3$  was enough to achieve this interesting sequential transformation. Molecular oxygen as an ideal green oxidant has attracted considerable attention in organic transformation because it is abundant, free, nontoxic, and free of contamination.<sup>13</sup> Moreover, the introduction of an amino group into 1,4-diketone skeletons made their further transformation more diverse. These inspired us to investigate this interesting transformation in depth.

The influence of solvent and base was investigated carefully (Table 2). Different solvents were evaluated by employing 20 mol % of  $K_2CO_3$  as the catalyst. Conducting the reaction in THF, DCE, or toluene only afforded a trace of 4aa (entries 2, 4,

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Table 2.	Screening	of the	Reaction	Conditions

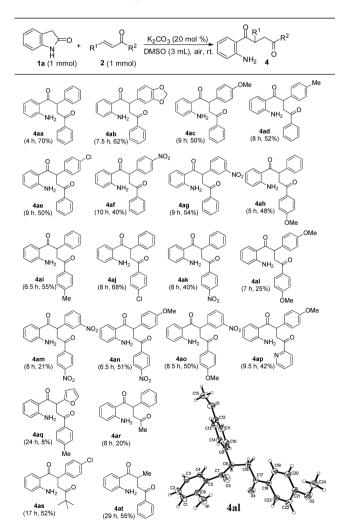
	0 H 1a (1 mmol) + Ph 2a (1 mmol)	conditions solvent (3 mL),		Ph O 4aa
entry	base (equiv)	solvent	time (h)	yield <sup>a</sup> (%)
1	$K_2CO_3$ (0.2)	CH <sub>3</sub> CN	24	17
2	$K_2CO_3$ (0.2)	THF	28	trace
3	$K_2CO_3$ (0.2)	acetone	20	11
4	$K_2 CO_3 (0.2)$	toluene	28	trace
5	$K_2CO_3$ (0.2)	DCE	28	trace
6	$K_2CO_3$ (0.2)	DMSO	4	70
7	$K_2CO_3(1)$	DMSO	2.5	67
8	$K_2CO_3$ (0.05)	DMSO	12	59
9	$K_2 CO_3 (0.2)$	DMF	12	69
10	$K_2 CO_3 (0.2)$	NMP	24	32
11 <sup>b</sup>	$K_2CO_3$ (0.2)	DMSO	24	4
12	NaOH (0.2)	DMSO	3	52
13	$Et_{3}N(0.2)$	DMSO	24	8
14	DMAP (0.2)	DMSO	24	9
15	piperidine (0.2)	DMSO	24	9
16	DBU (0.2)	DMSO	4	47

<sup>*a*</sup>Isolated yield. <sup>*b*</sup>Carried out under N<sub>2</sub> atomosphere.

and 5). Using NMP (N-methylpyrrolidone) as solvent gave 32% yield of 4aa (entry 10), and a much lower yield was obtained in acetonitrile and acetone (entries 1 and 3). Upon switching the solvent to DMSO, the yield of 4aa was dramatically improved to 70% (entry 6). Although DMF also gave a yield comparable to that of DMSO, the reaction took much longer to complete (entry 9). Further screening different bases for this transformation in DMSO revealed that organic bases such as Et<sub>3</sub>N, DMAP, and piperidine afforded very low yields of 4aa (entries 13-15) and the NaOH or DBU only gave moderate vields of 4aa (entries 12 and 16). The O<sub>2</sub> proved to be crucial to the reaction, and only 4% of 4aa could be obtained under N<sub>2</sub> atmosphere after 24 h (entry 11). Reducing the loading of K<sub>2</sub>CO<sub>3</sub> to 5 mol % led to longer reaction time and slightly lower yield (entry 8). Although employing 1 equiv of K<sub>2</sub>CO<sub>3</sub> could shorten the reaction time, no noticeable enhancement on the yield was obtained (entry 7).

With the optimal conditions in hand (Table 2, entry 6), the scope of the reaction was then investigated (Table 3). The  $R^1$  and  $R^2$  substituents of substrate 2 may be either electron-rich or electron-deficient aryl and alkyl groups. Most of the reactions gave moderate yields of products. When both of the aryl groups of  $R^1$  and  $R^2$  linked with strong electron-donating or strong electron-withdrawing groups, the yield was unsatisfactory (4al and 4am). When  $R^2$  was a methyl group, only 20% yield of

#### Table 3. Substrate Scope of the Enones

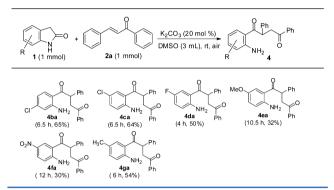


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product (4ar) was obtained. Replacing the methyl group with *tert*-butyl group increased the yield to 52% (4as). Pyridyl group was also tolerated in the transformation, producing the product 4ap in 42% yield. The furyl group gave a very low yield of product 4aq. The structure of product 4al was further established by single-crystal X-ray crystallographic analysis.

The applicability of other 2-oxindoles in this conversion was also evaluated by performing the reaction of them with chalcone 2a (Table 4). The results showed that an obvious

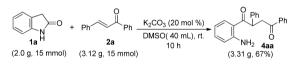
Table 4. Substrate Scope of the Oxindoles



electronic substrate effect existed. Either strong electrondonating or strong electron-withdrawing group substituted 2oxindole gave low yield of the product.

To show the practicality of this protocol, a large-scale experiment was carried out by performing the reaction of 1a (2.0 g, 15 mmol) with 3a (3.12 g, 15 mmol) in 40 mL of DMSO in the presence of 3 mmol of  $K_2CO_3$  (Scheme 2). In

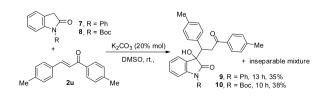




order to increase the concentration of oxygen in the solvent, air was simultaneously bubbled into the reaction mixture through a glass dropper. This transformation proceeded smoothly to afford **4aa** in 67% yield within 10 h.

Among the above investigated reactions, there was no substituent on the nitrogen atom of 2-oxindole. Next, the reactivity of *N*-substituted 2-oxindoles was considered (Scheme 3). For the reaction of *N*-phenyl-2-oxindole (7) with enone **2u** under the standard conditions, although the TLC indicated that full conversion to two almost equal amounts of products had occurred, the <sup>1</sup>H NMR analysis showed that both of them were not the anticipated product. The product with higher polarity was assigned as the hydroxylated Michael adduct **9** (35%)<sup>14</sup> but not the hydroperoxide. Because the characteristic chemical

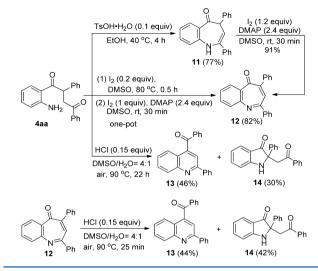
### Scheme 3. Reaction of *N*-Substituted 2-Oxindoles with Enone



shifts for -OOH at  $9-10 \text{ ppm}^{15}$  were not observed in the <sup>1</sup>H NMR spectrum of 9, the product with lower polarity was proved to be a mixture of three inseparable compounds. *N*-Boc-protected 2-oxindole (8) gave a result similar to that of *N*-phenyl-2-oxindole.

Due to the simultaneous existence of amino and carbonyl groups, the synthetic potential of the products 4 through the intramolecular reaction was investigated (Scheme 4). No

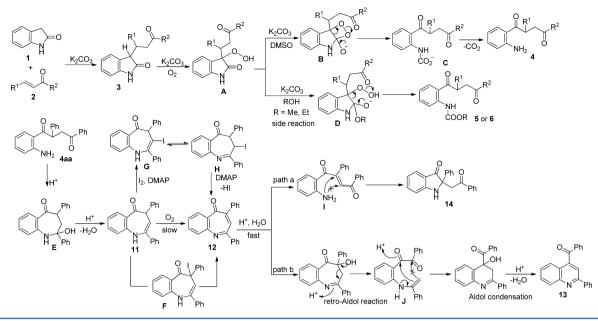
Scheme 4. Further Transformation of 4aa



reaction occurred by heating 4aa in ethanol with 20% mol of acetic acid or directly in acetic acid at reflux. When the HOAc was replaced by TsOH·H<sub>2</sub>O, the 4aa was smoothly converted to benzoazepinone 11 in 77% yield in ethanol at 40 °C. It seems that 4aa are liable to convert to 12 through a condensation-oxidation process. However, several oxidation conditions like Cu(OAc)<sub>2</sub>, PhI(OAc)<sub>2</sub>, DDQ, and K<sub>2</sub>S<sub>2</sub>O<sub>8</sub> could not realize this simple looking conversion. Finally, a concise route to access 12 through a two-step, one-pot reaction was explored. In the presence of 1.2 equiv of  $I_2$  and 2.4 equiv of DMAP, 11 could turn into the single product 12 in excellent vield. Surprisingly, the iodine could also catalyze the transformation of 4aa to 11 in DMSO at 80 °C, which could be explained by the formation of HI in the iodination catalyzing the conversion. Thus, a simple one-pot conversion of 4aa to 12 was realized. A mixture of 4aa and 0.2 equiv of I<sub>2</sub> was stirred in DMSO at 80 °C for 0.5 h. After being cooled to room temperature, 1.0 equiv of I2 and 2.4 equiv of DMAP were added, and the mixture was further stirred at room temperature for 30 min to give 12 in 82% yield. The structure of product 12 was unambiguously determined on the basis of single-crystal Xray diffraction analysis (see the Supporting Information). It was noteworthy that direct reaction of 4aa with 1.2 equiv of  $I_2$  and 2.4 equiv of DMAP led to a complex mixture with very low conversion. To the best our knowledge, the benzoazepinone skeleton like 11 and 12 was rather rare.<sup>16</sup> If 4aa was heated in a mixed solvent of  $DMSO/H_2O$  (4:1) in the presence of catalytic amounts of hydrochloric acid at 90 °C for 22 h, two interesting cyclization products 13<sup>17</sup> and 14 were provided in 46% and 30% yield, respectively. During the reaction, 11 was quickly formed as the intermediate. However, the conversion of 11 to 13 and 14 needed a very long reaction time.

A possible mechanism is depicted in Scheme 5. Initially, Michael addition of 2-oxindole to enone affords the Michael

#### Scheme 5. Proposed Mechanism



adduct 3. Under basic conditions, the reaction of 3 with O2 generates peroxide A.<sup>15a,18</sup> Intramolecular attacks of peroxide anion on the amide group produce the dioxetane intermediate B, which undergoes dissociation to generate C. Decarboxvlation of C formed the product 4. This type of cleavage of an oxindole ring under basic oxidative conditions has precedence in the literature.<sup>19</sup> However, a large excess amount of either NaH (3 equiv) or NaOH (>10 equiv) was needed, <sup>19a,b,d</sup> or a mixture was generated.<sup>19c,d</sup> If the reaction was carried out in methanol or ethanol, the competing intermolecular attack of alkoxy anion at the amide group provides the intermediate D, which undergoes sequential C-C and O-O bond cleavage to generate the side product 5 or 6. At present, we had no definite explanation for the generation of different products for Nprotected 2-oxindoles. It was possible that the substituent on the nitrogen atom was not benificial to the attack of the peroxide anion on the amide group. As a consequence, the oxidation of starting material by the generated peroxide led to the formation of hydroxylated product.

Under acidic conditions, the intramoleclar reaction of amino group with the carbonyl affords the enamine product 11. The iodination of 11 in the presence of DMAP provides the intermediate F. Another iodinated product G also could not be exluded.<sup>20</sup> Extrusion of HI from F or the imine H (equilibrates to G) by DMAP would afford the product 12. This indirect route (one-pot, two-step) could synthesize 12 in good yield within a short time. At high temperature, 11 is slowly oxidized by  $O_2$  to generate 12. Two possible reaction pathways exist for the rapid hydrolysis of 12 in the presence of H<sub>2</sub>O under acidic conditions. In path a, C=N bond cleavage gives the intermediate I, which undergoes selective intramolecular Michael addition to afford the product 14. In path b, the retro-Aldol condensation generates the intermediate J. The follow-up selective Aldol condensation produces the quinoline product 13. The water was essential to the hydrolysis of 12, if water was not added as the cosolvent, the hydrolysis was very slow. The hydrolysis of 12 was so fast that we could not observe the intermediate 12 on TLC during the hydrochloric acid-catalyzed reaction of 4aa in DMSO/H2O. In order to

prove this, the reaction of 12 in the presence of catalytic amount of hydrochloric acid was conducted in DMSO/H<sub>2</sub>O at 90 °C (Scheme 4). Full conversion of 12 was completed within 25 min along with the formation of 13 and 14 with the ratio of 1:1.

In summary, the  $K_2CO_3$ -catalyzed reaction of 2-oxindoles with enones for the preparation of a variety of 1,4-diketones bearing an amino group was disclosed. Simple experimental procedures and mild reaction conditions were used for the successive Michael addition—oxidation—ring-cleavage reactions. The further intramolecular reaction of the 1,4-diketone produced the benzoazepinone, quinoline, and 3-oxindole derivatives. Only atmospheric oxygen was needed in the oxidative process among these transformations.

#### EXPERIMENTAL SECTION

**General Information.** <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on 300 and 500 MHz (75 and 100 MHz for <sup>13</sup>C NMR) spectrometers. Melting points were determined on a micromelting point apparatus without corrections. Flash column chromatography was performed over silica gel (200–300 mesh).

**Preparation of Starting Materials.** 2-Oxindoles 1a–g, 7, and 2r are commercially available. N-Boc-2-oxindole 8 was prepared from 1a and  $(Boc)_2O$  according to a reported procedure.<sup>21</sup> Enones 2a–q,s were synthesized from the corresponding ketones and aldehydes catalyzed by NaOH.<sup>22</sup> (*E*)-1-Phenyl-2-buten-1-one 2t was prepared through the Friedel–Crafts reaction of benzene with (*E*)-2-butenoyl chloride.<sup>23</sup>

K<sub>2</sub>CO<sub>3</sub>-Catalyzed Reaction of 2-Oxindole (1a) with Chalcone (2a) in Ethanol or Methanol. A mixture of 2-oxindole (133 mg, 1 mmol), chalcone 2a (208 mg, 1 mmol), and K<sub>2</sub>CO<sub>3</sub> (28 mg, 0.2 mmol) was stirred in 3 mL of ethanol or methanol in a tube ( $\emptyset$  18 × 150 mm) under open air at room temperature for 48 or 24 h, respectively. Thirty milliliters of water was added, and the mixture was extracted with dichloromethane (20 mL × 3). The combined organic phases were dried over anhydrous sodium sulfate and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate/petroleum ether) to provide the corresponding products 5 (or 6), 4aa, and 3a + 3b.

**3a**: white solid; mp 129–130 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (d, J = 7.1 Hz, 2H), 7.65 (s, br, 1H), 7.57 (tt, J = 7.3, 1.3 Hz, 1H), 7.46 (t, J = 7.4 Hz, 2H), 7.31 (d, J = 7.3 Hz, 1H), 7.08–7.16 (m, 6H),

7.01 (td, *J* = 7.6, 1.0 Hz, 1H), 6.64 (d, *J* = 7.6 Hz, 1H), 4.17–4.29 (m, 2H), 3.88 (d, *J* = 3.4 Hz, 1H), 3.56 (dd, *J* = 20.6 Hz, 8.7 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  199.0, 178.4, 141.0, 139.7, 137.2, 133.3, 128.8, 128.5, 128.4, 128.3, 128.2, 128.0, 127.0, 124.9, 122.4, 109.3, 50.1, 42.0, 39.8; HRMS (ESI-Q-TOF) *m*/*z* [M + Na]<sup>+</sup> calcd for C<sub>23</sub>H<sub>19</sub>NNaO<sub>2</sub> 364.1313, found 364.1309.

**5**: yellow solid; 24.2 mg, 6%; mp 42–43 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  11.09 (s, 1H), 8.46 (dd, *J* = 8.6, 1.0 Hz, 1H), 8.10 (dd, *J* = 8.2, 1.5 Hz, 1H), 7.99 (d, *J* = 7.1 Hz, 2H), 7.58 (tt, *J* = 7.4, 1.3 Hz, 1H), 7.42–7.51 (m, 3H), 7.28–7.36 (m, 4H), 7.19–7.28 (m, 1H), 7.16–7.25 (m, 1H), 7.00 (ddd, *J* = 8.2, 7.4, 1.2 Hz, 1H), 5.37 (dd, *J* = 10.3, 3.5 Hz, 1H), 4.21 (dd, *J* = 18.2 Hz, 10.3 Hz, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 3.29 (dd, *J* = 18.2, 3.5 Hz, 1H), 1.30 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  202.6, 198.0, 154.0, 142.3, 138.8, 136.5, 134.9, 133.5, 131.6, 129.5, 128.8, 128.3, 128.1, 127.7, 121.4, 120.9, 119.3, 61.3, 49.5, 44.2, 14.6; HRMS (ESI-Q-TOF) *m*/*z* [M + Na]<sup>+</sup> calcd for C<sub>25</sub>H<sub>23</sub>NNaO<sub>4</sub> 424.1525, found 424.1521.

6: yellow solid; 37.9 mg, 10%; mp 97–99 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 11.13 (s, 1H), 8.44 (dd, J = 8.5, 0.8 Hz, 1H), 8.10 (dd, J = 8.2, 1.3 Hz, 1H), 7.98 (d, J = 7.1 Hz, 2H), 7.57 (tt, J = 7.4, 1.3 Hz, 1H), 7.20–7.36 (m, 5H), 7.00 (ddd, J = 8.2, 7.3, 1.1 Hz, 1H), 5.36 (dd, J = 10.4, 3.5 Hz, 1H), 4.21 (dd, J = 18.1, 10.4 Hz, 1H), 3.74 (s, 3H), 3.28 (dd, J = 18.1, 3.4 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 202.6, 198.0, 154.3, 142.1, 138.7, 136.3, 134.9, 133.5, 131.6, 129.4, 128.7, 128.3, 128.0, 127.6, 121.5, 120.9, 119.2, 52.3, 49.5, 44.2; HRMS (ESI-Q-TOF) m/z [M + Na]<sup>+</sup> calcd for C<sub>24</sub>H<sub>21</sub>NNaO<sub>4</sub> 410.1368, found 410.1366.

General Procedure for the  $K_2CO_3$ -Catalyzed Reaction of 2-Oxindoles (1) with Enones (2) in DMSO for the Preparation of 4. A mixture of 2-oxindoles 1 (1 mmol), enones 2 (1 mmol), and  $K_2CO_3$  (0.2 mmol) was stirred in 3 mL of DMSO in a tube ( $\emptyset$  18 × 150 mm) under open air at room temperature for a given time. Upon completion of the reaction as determined by TLC, 30 mL of water was added, and the mixture was extracted with dichloromethane (20 mL × 3). The combined organic phases were dried over anhydrous sodium sulfate and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate/petroleum ether) to provide the corresponding products 4.

**4aa:** yellow solid; 230 mg, 70%; mp 77–79 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.99 (d, J = 7.1 Hz, 2H), 7.91 (d, J = 7.7 Hz, 1H), 7.55 (tt, J = 7.3 Hz, 1.3 Hz, 1H), 7.44 (t, J = 7.5 Hz, 2H), 7.28–7.39 (m, 4H), 7.14–7.25 (m, 2H), 6.53–6.63 (m, 2H), 6.21 (s, br, 2H), 5.34 (dd, J = 10.0, 3.8 Hz, 1H), 4.19 (dd, J = 18.0, 10.0 Hz, 1H), 3.23 (dd, J = 17.9, 3.8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.7, 198.3, 151.2, 139.9, 136.7, 134.3, 133.3, 131.8, 129.2, 128.7, 128.3, 128.1, 127.2, 117.5, 116.0, 48.9, 43.9; HRMS (+ESI) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>20</sub>NO<sub>2</sub> 330.1494, found 330.1490.

**4ab**: yellow solid; 231 mg, 62%; mp 120–121 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.99 (d, J = 7.1 Hz, 2H), 7.91 (dd, J = 8.6, 1.4 Hz, 1H), 7.56 (tt, J = 7.3, 1.3 Hz, 1H), 7.44 (t, J = 7.5 Hz, 2H), 7.20 (ddd, J = 8.3, 7.3, 1.4 Hz, 1H), 6.84 (s, 1H), 6.82 (dd, J = 8.3, 1.9 Hz, 1H), 6.74 (dd, J = 7.5, 0.9 Hz, 1H), 6.55–6.63 (m, 2H), 6.20 (s, br, 2H), 5.92 (d, J = 1.4 Hz, 1H), 5.90 (d, J = 1.4 Hz, 1H), 5.26 (dd, J = 9.8, 4.0 Hz, 1H), 4.12 (dd, J = 18.0, 9.8 Hz, 1H), 3.21 (dd, J = 18.0, 3.9 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.7, 198.3, 151.2, 148.2, 146.8, 136.7, 134.3, 133.6, 133.3, 131.7, 128.7, 128.3, 121.4, 117.5, 117.4, 116.0, 108.9, 108.4, 101.2, 48.4, 43.9; HRMS (ESI) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>20</sub>NO<sub>4</sub> 374.1392, found 374.1384.

**4ac:** yellow solid; 180 mg, 50%; mp 135–136 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.99 (d, J = 7.1 Hz, 2H), 7.91 (dd, J = 8.6, 1.6 Hz, 1H), 7.55 (tt, J = 7.4, 1.3 Hz, 1H), 7.44 (t, J = 7.5 Hz, 2H), 7.27 (d, J = 8.9 Hz, 2H), 7.17 (ddd, J = 8.3, 7.3, 1.5 Hz, 1H), 6.84 (d, J = 8.9 Hz, 2H), 6.54–6.62 (m, 2H), 6.20 (s, br, 2H), 5.29 (dd, J = 9.8, 4.0 Hz, 1H), 4.14 (dd, J = 17.9, 9.8 Hz, 1H), 3.75 (s, 3H), 3.21 (dd, J = 17.9, 4.0 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.9, 198.4, 158.7, 151.2, 136.6, 134.2, 133.2, 131.8, 131.7, 129.1, 128.6, 128.2, 117.44, 117.35, 115.9, 114.5, 55.3, 47.9, 43.9; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>3</sub> 360.1600, found 360.1596.

**4ad**: yellow solid; 178 mg, 52%; mp 61–63 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (d, J = 7.1 Hz, 2H), 7.91 (dd, J = 8.5, 1.3 Hz,

1H), 7.52 (tt, *J* = 7.3, 1.3 Hz, 1H), 7.41 (t, *J* = 7.5 Hz, 2H), 7.23 (d, *J* = 8.0 Hz, 2H), 7.15 (ddd, J = 8.3, 7.2, 1.5 Hz, 1H), 7.10 (d, *J* = 8.0 Hz, 2H), 6.52–6.59 (m, 2H), 6.20 (s, br, 2H), 5.30 (dd, *J* = 10.0, 3.8 Hz, 1H), 4.16 (dd, *J* = 18.0, 10.0 Hz, 1H), 3.19 (dd, *J* = 18.0, 3.8 Hz, 1H), 2.27 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.8, 198.3, 151.1, 136.8, 136.7, 136.6, 134.1, 133.1, 131.7, 129.8, 128.6, 128.1, 127.8, 117.4, 117.3, 115.8, 48.4, 43.9, 21.0; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>2</sub> 344.1651, found 344.1642.

**4ae**: yellow solid; 182 mg, 50%; mp 99–101 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) *δ* 7.98 (d, *J* = 7.1 Hz, 2H), 7.86 (d, *J* = 8.2 Hz, 1H), 7.55 (tt, *J* = 7.3, 1.3 Hz, 1H), 7.44 (t, *J* = 7.5 Hz, 2H), 7.25–7.32 (m, 4H), 7.19 (ddd, *J* = 8.3, 7.3, 1.5 Hz, 1H), 6.54–6.62 (m, 2H), 6.22 (s, br, 2H), 5.33 (dd, *J* = 9.7, 4.1 Hz, 1H), 4.13 (dd, *J* = 18.0, 9.7 Hz, 1H), 3.22 (dd, *J* = 17.9, 4.1 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) *δ* 2002, 197.9, 151.2, 138.3, 136.4, 134.4, 133.3, 133.0, 131.5, 129.4, 129.2, 128.6, 128.1, 117.5, 117.0, 115.8, 47.9, 43.6; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>CINO<sub>2</sub> 364.1104, found 364.1097.

**4af:** yellow solid; 150 mg, 40%; mp 137–139 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.17 (d, J = 8.8 Hz, 2H), 7.98 (d, J = 7.1 Hz, 2H), 7.83 (dd, J = 8.3 Hz, 1.3 Hz, 1H), 7.52–7.61 (m, 3H), 7.46 (t, J = 7.5 Hz, 2H), 7.22 (ddd, J = 8.3, 7.3, 1.5 Hz, 1H), 6.56–6.65 (m, 2H), 6.26 (s, br, 2H), 5.48 (dd, J = 9.2 Hz, 4.4 Hz, 1H), 4.16 (dd, J = 17.9 Hz, 9.2 Hz, 1H), 3.30 (dd, J = 17.9 Hz, 4.5 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  199.3, 197.4, 151.5, 147.4, 147.1, 136.3, 134.9, 133.6, 131.3, 129.1, 128.8, 128.3, 124.4, 117.7, 116.9, 116.1, 48.4, 43.4; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>N<sub>2</sub>O<sub>4</sub> 375.1345, found 375.1336.

**4ag**: yellow solid; 202 mg, 54%; mp 141–142 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) *δ* 8.26 (t, *J* = 1.9 Hz, 1H), 8.09 (ddd, *J* = 8.2, 2.2, 1.0 Hz, 1H), 7.99 (d, *J* = 7.1 Hz, 2H), 7.87 (dd, *J* = 8.7, 1.5 Hz, 1H), 7.73 (dt, *J* = 7.7, 1.3 Hz, 1H), 7.58 (tt, *J* = 7.3, 1.3 Hz, 1H), 7.42–7.53 (m, 3H), 7.23 (ddd, *J* = 8.5, 7.2, 1.4 Hz, 1H), 6.58–6.66 (m, 2H), 6.27 (s, br, 2H), 5.49 (dd, *J* = 9.3, 4.5 Hz, 1H), 4.18 (dd, *J* = 18.0, 9.3 Hz, 1H), 3.33 (dd, *J* = 18.0, 4.5 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) *δ* 199.4, 197.5, 151.5, 148.7, 141.9, 136.3, 134.8, 134.5, 133.6, 131.3, 130.1, 128.8, 128.2, 123.1, 122.4, 117.7, 116.8, 116.1, 47.9, 43.5; HRMS (ESI-Q-TOF) *m/z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>N<sub>2</sub>O<sub>4</sub> 375.1345, found 375.1341.

**4ah**: yellow solid; 172 mg, 48%; mp 77–78 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (d, J = 9.0 Hz, 2H), 7.91(d, J = 8.0 Hz, 1H), 7.27–7.39 (m, 4H), 7.13–7.24 (m, 2H), 6.90 (d, J = 9.0 Hz, 2H), 6.53–6.61 (m, 2H), 6.21 (s, br, 2H), 5.33 (dd, J = 10.0, 3.8 Hz, 1H), 4.13 (dd, J = 17.8, 10.0 Hz, 1H), 3.85 (s, 3H), 3.19 (dd, J = 17.8, 3.8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.8, 196.8, 163.6, 151.2, 140.0, 134.2, 131.7, 130.5, 129.7, 129.1, 128.0, 127.1, 117.45, 117.41, 115.9, 113.7, 55.5, 48.8, 43.6; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>3</sub> 360.1600, found 360.1592.

**4ai:** yellow solid; 189 mg, 55%; mp 69–71 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.86–7.94 (m, 3H), 7.27–7.38 (m, 4H), 7.13–7.25 (m, 4H), 6.53–6.62 (m, 2H), 6.21 (s, br, 2H), 5.33 (dd, *J* = 10.0, 3.8 Hz, 1H), 4.16 (dd, *J* = 17.9, 10.0 Hz, 1H), 3.21 (dd, *J* = 17.9, 3.8 Hz, 1H), 2.39 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.7, 197.8, 151.2, 143.9, 139.9, 134.13, 134.10, 131.7, 129.2, 129.1, 128.3, 128.0, 127.1, 117.4, 117.3, 115.7, 48.7, 43.7, 21.6; HRMS (ESI-Q-TOF) *m/z* [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>2</sub> 344.1651, found 344.1647.

**4aj**: yellow solid; 247 mg, 68%; mp 115–116 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) *δ* 7.85–7.92 (m, 3H), 7.37 (d, *J* = 8.6 Hz, 2H), 7.24–7.35 (m, 4H), 7.09–7.24 (m, 2H), 6.49–6.58 (m, 2H), 6.22 (s, br, 2H), 5.31 (dd, *J* = 10.0, 3.7 Hz, 1H), 4.12 (dd, *J* = 17.9, 10.1 Hz, 1H), 3.15 (dd, *J* = 18.0, 3.8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) *δ* 200.5, 197.1, 151.2, 139.8, 139.6, 135.0, 134.3, 131.7, 129.7, 129.2, 128.9, 128.0, 127.3, 117.5, 117.3, 115.9, 48.9, 43.8; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>ClNO<sub>2</sub> 364.1104, found 364.1095.

**4ak**: yellow solid; 150 mg, 40%; mp 135–136 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.30 (d, J = 9.0 Hz, 2H), 8.14 (d, J = 9.0 Hz, 2H), 7.87 (dd, J = 8.2 Hz, 1.1 Hz, 1H), 7.29–7.38 (m, 4H), 7.16–7.27 (m, 2H), 6.53–6.64 (m, 2H), 6.23 (s, br, 2H), 5.34 (dd, J = 10.1, 3.7 Hz, 1H), 4.19 (dd, J = 18.0, 10.1 Hz, 1H), 3.21 (dd, J = 17.9, 3.8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.2, 197.1, 151.3, 150.4, 141.1, 139.5, 134.5, 131.8, 129.4, 129.3, 128.0, 127.5, 123.9, 117.5, 117.1, 116.0,

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49.2, 44.3; HRMS (ESI-Q-TOF)  $m/z [M + H]^+$  calcd for  $C_{22}H_{19}N_2O_4$  375.1345, found 375.1338.

**4al**: yellow solid; 97 mg, 25%; mp 132 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (d, *J* = 9.0 Hz, 2H), 7.92 (dd, *J* = 8.7 Hz, 1.5 Hz, 1H), 7.27 (d, *J* = 8.9 Hz, 2H), 7.17 (ddd, *J* = 8.3, 7.3, 1.4 Hz, 1H), 6.91 (d, *J* = 9.0 Hz, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 6.54–6.61 (m, 2H), 6.20 (s, br, 2H), 5.28 (dd, *J* = 9.8, 4.0 Hz, 1H), 4.09 (dd, *J* = 17.7, 9.8 Hz, 1H), 3.85 (s, 3H), 3.75 (s, 3H), 3.17 (dd, *J* = 17.7, 4.0 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  201.2, 197.0, 163.6, 158.7, 151.2, 134.2, 132.0, 131.8, 130.5, 129.9, 129.1, 117.6, 117.5, 116.0, 114.6, 113.8, 55.6, 55.3, 48.0, 43.6; HRMS (ESI-Q-TOF) *m/z* [M + Na]<sup>+</sup> calcd for C<sub>24</sub>H<sub>23</sub>NNaO<sub>4</sub> 412.1525, found 412.1516.

**4am**: yellow solid; 88 mg, 21%; mp 78–79 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.32 (d, *J* = 9.0 Hz, 2H), 8.26 (t, *J* = 2.0 Hz, 1H), 8.16 (d, *J* = 9.0 Hz, 2H), 8.12 (ddd, *J* = 8.2, 2.2, 1.0 Hz, 1H), 7.82 (dd, *J* = 8.2, 1.1 Hz, 1H), 7.72 (dt, *J* = 7.9, 1.3 Hz, 1H), 7.52 (t, *J* = 8.0 Hz, 1H), 7.24 (ddd, *J* = 8.4 Hz, 7.1 Hz, 1.4 Hz, 1H), 6.58–6.66 (m, 2H), 6.29 (s, br, 2H), 5.48 (dd, *J* = 9.7, 4.2 Hz, 1H), 4.23 (dd, *J* = 18.0, 9.6 Hz, 1H), 3.29 (dd, *J* = 18.0, 4.1 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  198.9, 196.3, 151.6, 150.5, 148.8, 141.5, 140.7, 135.0, 134.3, 131.3, 130.3, 129.3, 124.0, 123.0, 122.6, 117.8, 116.4, 116.2, 48.2, 43.9; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>18</sub>N<sub>3</sub>O<sub>6</sub> 420.1196, found 420.1189.

**4an**: yellow solid; 206 mg, 51%; mp 177–178 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.30 (d, J = 9.0 Hz, 2H), 8.14 (d, J = 9.0 Hz, 2H), 7.87 (dd, J = 8.2 Hz, 1.1 Hz, 1H), 7.26 (d, J = 8.9 Hz, 2H), 7.20 (ddd, J = 8.3, 7.0, 1.5 Hz, 1H), 6.85 (d, J = 8.8 Hz, 2H), 6.55–6.63 (m, 2H), 6.21 (s, br, 2H), 5.29 (dd, J = 10.0, 3.9 Hz, 1H), 4.15 (dd, J = 18.0, 10.0 Hz, 1H), 3.76 (s, 3H), 3.19 (dd, J = 17.9, 3.9 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.5, 197.2, 158.9, 151.3, 150.4, 141.2, 134.5, 131.8, 131.4, 129.3, 129.0, 123.9, 117.5, 117.1, 116.0, 114.7, 55.4, 48.3, 44.3; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>21</sub>N<sub>2</sub>O<sub>5</sub> 405.1450, found 405.1440.

**4ao:** yellow solid; 202 mg, 50%; mp 158 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.26 (t, *J* = 1.9 Hz, 1H), 8.08 (ddd, *J* = 8.2, 2.2, 1.0 Hz, 1H), 7.97 (d, *J* = 9.0 Hz, 2H), 7.88 (dd, *J* = 8.6 Hz, 1.3 Hz, 1H), 7.73 (dt, *J* = 7.8 Hz, 1.3 Hz, 1H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.22 (ddd, *J* = 8.3, 7.1, 1.4 Hz, 1H), 6.93 (d, *J* = 8.9 Hz, 2H), 6.58–6.65 (m, 2H), 6.27 (s, br, 2H), 5.48 (dd, *J* = 9.2, 4.5 Hz, 1H), 4.12 (dd, *J* = 17.8, 9.3 Hz, 1H), 3.87 (s, 3H), 3.29 (dd, *J* = 17.7, 4.6 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  199.6, 195.9, 163.8, 151.5, 148.7, 142.0, 134.8, 134.5, 131.3, 130.6, 130.0, 129.4, 123.1, 122.4, 117.7, 116.9, 116.1, 113.9, 55.6, 48.0, 43.2; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>21</sub>N<sub>2</sub>O<sub>5</sub> 405.1450, found 405.1442.

**4ap**: yellow solid; 151 mg, 42%; mp 118–120 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.68 (d, J = 4.4 Hz, 1H), 8.00 (d, J = 7.8 Hz, 1H), 7.93 (dd, J = 8.7 Hz, 1.4 Hz, 1H), 7.80 (td, J = 7.7 Hz, 1.7 Hz, 1H), 7.45 (ddd, J = 7.5 Hz, 4.8 Hz, 1.2 Hz, 1H), 7.27 (d, J = 8.7 Hz, 2H), 7.18 (ddd, J = 8.4, 7.3, 1.4 Hz, 1H), 6.82 (d, J = 8.7 Hz, 2H), 6.55–6.63 (m, 2H), 6.20 (s, br, 2H), 5.27 (dd, J = 10.1, 4.0 Hz, 1H), 4.41 (dd, J = 18.8, 10.2 Hz, 1H), 3.75 (s, 3H), 3.47 (dd, J = 18.8, 4.0 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  201.1, 200.1, 158.6, 153.1, 151.1, 149.1, 136.9, 134.1, 131.9, 131.8, 129.2, 127.3, 121.9, 117.4, 115.8, 114.4, 55.3, 48.0, 43.2; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub> 361.1552, found 361.1544.

**4aq:** yellow solid; 27 mg, 8%; mp 69 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.99 (dd, *J* = 8.2, 1.3 Hz, 1H), 7.91 (d, *J* = 8.2 Hz, 2H), 7.33 (dd, *J* = 1.8, 0.8 Hz, 1H), 7.21–7.28 (m, 3H), 6.60–6.69 (m, 2H), 6.28 (dd, *J* = 3.2, 1.9 Hz, 1H), 6.20 (s, br, 2H), 6.15 (d, *J* = 3.2 Hz, 1H), 5.48 (dd, *J* = 9.7, 4.1 Hz, 1H), 4.14 (dd, *J* = 17.9, 9.7 Hz, 1H), 3.39 (dd, *J* = 17.9, 4.1 Hz, 1H), 2.41 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  198.3, 197.5, 152.7, 151.3, 144.2, 142.1, 134.6, 134.1, 131.7, 129.4, 128.4, 117.5, 117.2, 116.1, 110.8, 107.1, 42.2, 40.4, 21.8; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>20</sub>NO<sub>3</sub> 334.1443, found 334.1433.

**4ar**: yellow oil; 53 mg, 20%; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.80 (dd, *J* = 8.2 Hz, 1.4 Hz, 1H), 7.23–7.30 (m, 4H), 7.12–7.22 (m, 2H), 6.58 (dd, J = 8.3, 0.9 Hz, 1H), 6.53 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 6.22 (s, br, 2H), 5.12 (dd, *J* = 9.9, 4.2 Hz, 1H), 3.55 (dd, *J* = 17.8, 9.8 Hz, 1H), 2.71 (dd, *J* = 17.8, 4.2 Hz, 1H), 2.18 (s, 3H); <sup>13</sup>C NMR (75

MHz, CDCl<sub>3</sub>)  $\delta$  207.2, 200.7, 151.2, 139.9, 134.3, 131.8, 129.2, 127.9, 127.2, 117.4, 117.3, 115.9, 49.0, 48.3, 30.2; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>NO<sub>2</sub> 268.1338, found 268.1330.

**4as**: yellow solid; 178 mg, 52%; mp 99–101 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.80 (d, *J* = 8.2 Hz, 1H), 7.18–7.25 (m, 4H), 7.13 (t, *J* = 7.5 Hz, 1H), 6.53 (t, *J* = 7.6 Hz, 2H), 6.23 (s, br, 2H), 5.13 (dd, *J* = 9.5, 4.4 Hz, 1H), 3.59 (dd, *J* = 18.0, 9.6 Hz, 1H), 2.76 (dd, *J* = 18.0, 4.4 Hz, 1H), 1.13 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  214.0, 200.3, 151.1, 138.4, 134.3, 132.8, 131.4, 129.3, 129.1, 117.4, 117.0, 115.8, 47.7, 43.9, 41.9, 26.4; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>23</sub>ClNO<sub>2</sub> 344.1417, found 344.1406.

**4at:** yellow solid; 147 mg, 55%; mp 76–77 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.00 (d, J = 7.1 Hz, 2H), 7.91 (dd, J = 8.2, 1.1 Hz, 1H), 7.55 (tt, J = 7.3, 1.3 Hz, 1H), 7.45 (t, J = 7.4 Hz, 2H), 7.26 (ddd, J = 8.4, 7.1, 1.4 Hz, 1H), 6.62–6.72 (m, 2H), 6.22 (s, br, 2H), 4.12–4.28 (m, 1H), 3.69 (dd, J = 18.0, 8.1 Hz, 1H), 3.06 (dd, J = 18.0, 5.2 Hz, 1H), 1.29 (d, J = 7.2 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  205.4, 198.7, 151.1, 136.8, 134.3, 133.2, 131.2, 128.6, 128.2, 117.6, 116.8, 116.0, 42.4, 36.4, 18.6; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>NO<sub>2</sub> 268.1338, found 268.1330.

**4ba**: yellow solid; 236 mg, 65%; mp 59–60 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.98 (d, J = 7.1 Hz, 2H), 7.82 (d, J = 8.8 Hz, 1H), 7.55 (tt, J = 7.3, 1.3 Hz, 1H), 7.43 (t, J = 7.4 Hz, 2H), 7.19–7.35 (m, SH), 6.57 (d, J = 2.0 Hz, 1H), 6.52 (dd, J = 8.7 Hz, 2.0 Hz, 1H), 6.31 (s, br, 2H), 5.25 (dd, J = 10.2, 3.7 Hz, 1H), 4.18 (dd, J = 18.0, 10.2 Hz, 1H), 3.21 (dd, J = 18.0, 3.6 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.0, 198.3, 152.0, 140.1, 139.6, 136.5, 133.4, 133.1, 129.3, 128.7, 128.2, 128.0, 127.4, 116.6, 116.4, 115.9, 49.0, 43.9; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>ClNO<sub>2</sub> 364.1104, found 364.1095.

**4ca**: yellow solid; 232 mg, 64%; mp 126–127 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) *δ* 7.99 (d, *J* = 7.3 Hz, 2H), 7.87 (d, *J* = 2.3 Hz, 1H), 7.56 (t, *J* = 7.3 Hz, 1H), 7.45 (t, *J* = 7.5 Hz, 2H), 7.21–7.38 (m, 5H), 7.13 (dd, *J* = 8.8 Hz, 2.3 Hz, 1H), 6.55 (d, *J* = 8.9 Hz, 1H), 6.21 (s, br, 2H), 5.24 (dd, *J* = 10.3, 3.5 Hz, 1H), 4.19 (dd, *J* = 18.0, 10.3 Hz, 1H), 3.24 (dd, *J* = 18.0, 3.5 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) *δ* 199.8, 198.2, 149.7, 139.2, 136.5, 134.3, 133.4, 130.8, 129.4, 128.7, 128.3, 128.0, 127.5, 120.2, 118.9, 118.0, 48.9, 44.0; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>CINO<sub>2</sub> 364.1104, found 364.1101.

**4da**: yellow solid; 174 mg, 50%; mp 104 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.98 (d, *J* = 7.1 Hz, 2H), 7.52–7.61 (m, 2H), 7.44 (t, *J* = 7.5 Hz, 2H), 7.20–7.38 (m, 5H), 6.96 (ddd, *J* = 9.0, 7.7, 2.9 Hz, 1H), 6.55 (dd, *J* = 9.1, 4.7 Hz, 1H), 6.07 (s, br, 2H), 5.21 (dd, *J* = 10.3, 3.5 Hz, 1H), 4.19 (dd, *J* = 18.0, 10.3 Hz, 1H), 3.23 (dd, *J* = 18.0, 3.5 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  199.8 (d, *J*<sub>4,C-F</sub> = 2.8 Hz), 198.2, 153.3 (d, *J*<sub>1,C-F</sub> = 234.2 Hz), 147.7, 139.2, 136.4, 133.3, 129.3, 128.6, 128.2, 127.9, 127.4, 122.4 (d, *J*<sub>2,C-F</sub> = 23.6 Hz), 118.6 (d, *J*<sub>3,C-F</sub> = 7.0 Hz), 116.7 (d, *J*<sub>4,C-F</sub> = 5.5 Hz), 116.1 (d, *J*<sub>2,C-F</sub> = 22.4 Hz), 48.9, 44.0; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>FNO<sub>2</sub> 348.1400, found 348.1392.

**4ea**: yellow solid; 116 mg, 32%; mp 88–90 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) *δ* 7.99 (d, *J* = 7.1 Hz, 2H), 7.55 (tt, *J* = 7.3, 1.3 Hz, 1H), 7.44 (t, *J* = 7.4 Hz, 2H), 7.29–7.39 (m, 5H), 7.18–7.25 (m, 1H), 6.87 (dd, *J* = 9.0, 2.9 Hz, 1H), 6.56 (d, 9.0 Hz, 1H), 5.93(s, br, 2H), 5.29 (dd, *J* = 9.9, 3.8 Hz, 1H), 4.18 (dd, *J* = 18.0, 9.9 Hz, 1H), 3.66 (s, 3 H), 3.22 (dd, *J* = 17.9, 3.8 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) *δ* 200.1, 198.3, 150.0, 146.0, 140.0, 136.7, 133.3, 129.3, 128.7, 128.3, 128.0, 127.3, 123.6, 118.9, 117.1, 113.8, 55.8, 49.4, 43.9; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>3</sub> 360.1600, found 360.1593.

**4fa**: yellow solid; 112 mg, 30%; mp 196–197 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.99 (d, *J* = 2.5 Hz, 1H), 7.96–8.06 (m, 3H), 7.58 (tt, *J* = 7.4, 1.3 Hz, 1H), 7.46 (t, *J* = 7.5 Hz, 2H), 8.03 (dd, *J* = 9.2, 2.5 Hz, 1H), 7.99 (d, *J* = 7.1 Hz, 2H), 7.23–7.31 (m, 1H), 6.98 (s, br, 2H), 6.59 (d, *J* = 9.2 Hz, 1H), 5.33 (dd, *J* = 10.7, 3.2 Hz, 1H), 4.23 (dd, *J* = 18.2 Hz, 10.7 Hz, 1H), 3.30 (dd, *J* = 18.1 Hz, 3.2 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ200.2, 198.2, 155.5, 138.5, 136.9, 136.3, 133.6, 129.6, 129.4, 129.2, 128.8, 128.3, 128.0, 127.8, 117.3, 115.3, 49.0, 44.0; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>19</sub>N<sub>2</sub>O<sub>4</sub> 375.1345, found 375.1340.

**4ga:** yellow solid; 184 mg, 54%; mp 132 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.98 (d, *J* = 7.1 Hz, 2H), 7.71 (s, 1H), 7.52 (t, *J* = 7.3 Hz, 1H), 7.41 (t, *J* = 7.5 Hz, 2H), 7.36 (t, *J* = 7.1 Hz, 2H), 7.29 (t, *J* = 7.5 Hz, 2H), 7.20 (t, *J* = 7.1 Hz, 1H), 6.99 (dd, *J* = 8.4, 1.7 Hz, 1H), 6.50 (d, *J* = 8.4 Hz, 1H), 6.05 (s, br, 2H), 5.35 (dd, *J* = 9.9, 3.8 Hz, 1H), 4.18 (dd, *J* = 17.9, 9.9 Hz, 1H), 3.22 (dd, *J* = 18.0, 3.9 Hz, 1H), 2.17 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 200.4, 198.3, 149.1, 140.0, 136.6, 135.4, 133.1, 131.2, 129.1, 128.5, 128.1, 128.0, 127.1, 124.5, 117.5, 117.2, 48.6, 43.8, 20.5; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>NO<sub>2</sub> 344.1651, found 344.1642.

 $K_2CO_3$ -Catalyzed Reactions of *N*-Phenyl-2-oxindole (7) or *N*-Boc-2-Oxindole (8) with Enone 2u. A mixture of *N*-phenyl-2oxindole 7 or *N*-phenyl-2-oxindole 8 (1 mmol), enone 2u (1 mmol), and  $K_2CO_3$  (0.2 mmol) was stirred in 4 mL of DMSO at room temperature under open-air conditions for given time. After completion of the reaction determined by TLC, 30 mL of water was added and the mixture was extracted with dichloromethane (20 mL × 3). The combined organic phases were dried over anhydrous sodium sulfate and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate/petroleum ether) to provide two fractions. The first fraction with low polarity was mixture of three inseparable compounds, and the second fraction with high polarity corresponds to product 9 (162 mg, 35%) or 10 (186 mg, 38%).

9: white solid; mp 201–203 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ 7.89 (d, *J* = 8.2 Hz, 2H), 7.59 (dd, *J* = 7.2, 1.2 Hz, 1H), 7.30–7.40 (m, 3H), 7.21–7.29 (m, 3H), 7.18 (td, *J* = 7.5, 1.4 Hz, 1H), 6.84–6.92 (m, 2H), 6.69–6.77 (m, 4H), 6.49 (d, *J* = 7.7 Hz, 1H), 4.00–4.13 (m, 2H), 3.52–3.68 (m, 2H), 2.41 (s, 3H), 2.23 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  198.6, 177.2, 144.2, 144.1, 137.0, 134.49, 134.47, 133.6, 129.8, 129.5, 129.4, 129.0, 128.6, 128.4, 128.2, 127.8, 126.5, 125.1, 123.1, 109.6, 79.7, 49.2, 38.5, 21.7, 21.0; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>31</sub>H<sub>28</sub>NO<sub>3</sub> 462.2069, found 462.2059.

**10**: white solid; mp 77–79 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.83 (d, *J* = 8.2 Hz, 2H), 7.59 (d, *J* = 7.9 Hz, 1H), 7.41 (dd, *J* = 7.4, 1.0 Hz, 1H), 7.32 (td, *J* = 7.7, 1.4 Hz, 1H), 7.17–7.25 (m, 3H), 6.85 (d, *J* = 7.9 Hz, 2H), 6.66 (d, *J* = 8.1 Hz, 2H), 3.79–4.01 (m, 3H), 3.47–3.61 (m, 1H), 2.39 (s, 3H), 2.18 (s, 3H), 1.48 (s, 9H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  198.4, 177.0, 148.2, 144.0, 139.8, 136.8, 134.3, 133.7, 129.9, 129.3, 128.7, 128.5, 128.3, 127.0, 124.6, 124.3, 115.0, 83.9, 79.1, 49.4, 38.0, 27.9, 21.7, 21.0; HRMS (ESI-Q-TOF) *m*/*z* [M + Na]<sup>+</sup> calcd for C<sub>30</sub>H<sub>31</sub>NNaO<sub>5</sub> 508.2100, found 508.2092.

TsOH·H<sub>2</sub>O-Catalyzed-Intramolecular Reaction of 4aa for the Synthesis of 11. A mixture of 4aa (53 mg, 0.16 mmol) and TsOH· $H_2O$  (3.1 mg, 0.016 mmol) in ethanol (0.4 mL) were stirred at 40 °C for 4 h until the disappearance of 4aa as determined by TLC. A saturated aqueous solution of NaHCO<sub>3</sub> (15 mL) was added, and the mixture was extracted with dichloromethane (15 mL × 3). The combined organic layers was dried over MgSO<sub>4</sub> and concentrated in vacuo. Purification on silica gel (ethyl acetate/petroleum ether) afforded the product 11 (38.5 mg, 77%).

11: yellow solid; mp 80–81 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ 8.00 (dd, J = 8.0, 1.6 Hz, 1H), 7.45–7.51 (m, 2H), 7.27–7.45 (m, 9H), 7.10 (d, J = 8.2 Hz, 1H), 6.99 (ddd, J = 8.0, 7.1, 0.9 Hz, 1H), 6.66 (s, br, 1H), 5.29 (dd, J = 7.0, 1.7 Hz, 1H), 4.29 (d, J = 7.0 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  185.2, 144.3, 141.5, 137.3, 137.1, 132.7, 131.2, 129.6, 129.4, 129.0, 128.5, 127.3, 125.4, 119.5, 118.3, 105.7, 57.4; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>18</sub>NO 312.1388, found 312.1380.

**One-Pot Two-Step Conversion of 4aa to 12.** A mixture of 4aa (36 mg, 0.11 mmol) and  $I_2$  (5.5 mg, 0.02 mmol) was stirred in 0.4 mL of DMSO at 80 °C for 30 min until the disappearance of 4aa as determined by TLC. After the mixture was cooled to room temperature,  $I_2$  (27 mg, 0.11 mmol) and DMAP (32 mg, 0.26 mmol) were added, and the mixture was stirred at room temperature for 30 min. Upon completion of the reaction, the mixture was quenched with saturated aqueous solution of  $Na_2S_2O_3$  and then extracted with dichloromethane (3 × 15 mL). The organic extracts were dried over sodium sulfate and concentrated in vacuo. The residue

was purified by column chromatography on silica gel (ethyl acetate/ petroleum ether) to provide the product **12** (27.8 mg, 82%).

12: yellow solid; mp 250 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  8.03– 8.10 (m, 2H), 7.94 (dd, J = 7.9, 1.5 Hz, 1H), 7.89 (dd, J = 8.0, 0.9 Hz, 1H), 7.75 (ddd, J = 8.1, 7.2, 1.6 Hz, 1H), 7.65–7.72 (m, 2H), 7.49– 7.56 (m, 4H), 7.48 (s, 1H), 7.40–7.47 (m, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  192.3, 159.9, 151.6, 146.1, 140.8, 137.2, 135.1, 132.9, 131.4, 130.7, 129.6, 128.9, 128.8, 128.8, 128.6, 128.3, 128.2, 125.8; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>16</sub>NO 310.1232, found 310.1224.

Hydrochloric Acid Catalyzed Transformation of 4aa to 13 and 14 in DMSO/H<sub>2</sub>O. Hydrochloric acid (12 mol/L, 2.5  $\mu$ L, 0.03 mmol) was added to a solution of 4aa (65.5 mg, 0.2 mmol) in DMSO/H<sub>2</sub>O (0.4 mL/0.1 mL). The mixture was stirred at 90 °C under open air conditions for 22 h. After completion of the reaction as determined by TLC, the mixture was quenched with saturated NaHCO<sub>3</sub> and then extracted with dichloromethane (20 mL × 3). The combined organic layers were dried over anhydrous sodium sulfate, filtered, and concentrated to give a residue, which was purified by column chromatography on silica gel (ethyl acetate/petroleum ether) to afford product 13 (28.4 mg, 46%, lower polarity) and product 14 (19.4 mg, 30%, higher polarity. The compound 14 has same polarity with 4aa in the eluent of ethyl acetate/petroleum ether. However, they can be separated on TLC using isopropyl ether/n-hexane = 5:4 as the eluent).

13: yellow solid; mp 105–106 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.27 (ddd, J = 8.5, 1.1, 0.6 Hz, 1H), 8.13–8.19 (m, 2H), 7.89–7.94 (m, 2H), 7.88 (s, 1H), 7.83–7.89 (m, 1H), 7.78 (ddd, J = 8.4, 6.9, 1.4 Hz, 1H), 7.66 (tt, J = 7.4, 1.3 Hz, 1H), 7.44–7.57 (m, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 196.4, 156.6, 148.8, 145.4, 139.0, 136.7, 134.3, 130.5, 130.35, 130.33, 129.8, 129.0, 128.9, 127.6, 127.4, 125.2, 124.0, 117.6; HRMS (ESI-Q-TOF) m/z [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>16</sub>NO 310.1232, found 310.1223.

14: yellow solid; mp 150 °C; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.92 (d, *J* = 7.1 Hz, 2H), 7.52–7.61 (m, 4H), 7.40–7.52 (m, 3H), 7.28 (tt, *J* = 6.9, 1.5 Hz, 2H), 7.21 (tt, *J* = 7.2, 1.3 Hz, 1H), 6.96 (d, *J* = 8.3 Hz, 1H), 6.81 (ddd, *J* = 7.8, 7.2, 0.7 Hz, 1H), 6.31 (s, br, 1H), 4.45 (d, *J* = 18.0 Hz, 1H), 3.18 (d, *J* = 17.9 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  200.8, 197.9, 160.4, 138.1, 137.9, 136.7, 133.9, 128.9, 128.8, 128.2, 127.7, 125.8, 125.4, 119.0, 118.2, 111.9, 69.4, 44.8; HRMS (ESI-Q-TOF) *m*/*z* [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>18</sub>NO<sub>2</sub> 328.1338, found 328.1326.

Conversion of 12 to 13 and 14 with a Catalytic Amount of Hydrochloric Acid in DMSO/H<sub>2</sub>O. Hydrochloric acid (12 mol/L,  $2.5 \,\mu$ L, 0.03 mmol) was added to a solution of 12 (55 mg, 0.18 mmol) in DMSO/H<sub>2</sub>O (0.4 mL/0.1 mL). The mixture was stirred at 90 °C until completion of the reaction as determined by TLC (within 25 min). The resulting mixture was quenched with saturated aqueous NaHCO<sub>3</sub> and extracted with dichloromethane (3 × 15 mL). The organic extracts were dried over sodium sulfate, filtered, and concentrated to give a residue that was purified by column chromatography on silica gel (ethyl acetate/petroleum ether) to afford product 13 (24.4 mg, 44%) and product 14 (24.7 mg, 42%).

#### ASSOCIATED CONTENT

#### **S** Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b02054.

<sup>1</sup>H and <sup>13</sup>C NMR spectra of the products (PDF) X-ray crystallographic data of **3a** (CIF) X-ray crystallographic data of **4al**(CIF) X-ray crystallographic data of **12**(CIF)

#### AUTHOR INFORMATION

#### **Corresponding Author**

\*E-mail: estally@yahoo.com

#### Notes

The authors declare no competing financial interest.

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#### REFERENCES

(1) For books, see: (a) Li, J. J.; Corey, E. J. Name Reactions in Heterocyclic Chemistry; Wiley-VCH: Weinheim, 2005. (b) Comprehensive Heterocyclic Chemistry III; Katritzky, A. R., Ramsden, C. A., Scriven, E. F., Taylor, R. J. K., Eds.; Elsevier: Oxford, 2008; Bergman, J.; Janosik, T. Vol. 3, pp 269–351; Graening, T.; Thrun, F. Vol. 3, pp 498–569; Sato, O.; Nakayama, J. Vol. 3, pp 844–930. (c) Ghosh, U.; Ganessunker, D.; Sattigeri, V. J.; Carlson, K. E.; Mortensen, D. J.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. Bioorg. Med. Chem. 2003, 11, 629. (d) Gravel, M.; Holmes, J. M. In Comprehensive Organic Synthesis II; Knochel, P., Molander, G. A., Eds.; Elsevier: Oxford, U.K., 2014; Vol. 4, pp 1384.

(2) (a) Dzvinchuk, I. B.; Nesterenko, A. M.; Lozinskii, M. O. Chem. Heterocycl. Compd. 2008, 44, 190. (b) Ahmed, S. A.; Khairou, K. S.; Asghar, B. H.; Muathen, H. A.; Nahas, N. M. A.; Alshareef, H. F. Tetrahedron Lett. 2014, 55, 2190. (c) Ghosh, U.; Ganessunker, D.; Sattigeri, V. J.; Carlson, K. E.; Mortensen, D. J.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. Bioorg. Med. Chem. 2003, 11, 629.

(3) (a) Clive, D. J.; Wang, J. J. Org. Chem. 2004, 69, 2773. (b) Chen, Z.-H.; Chen, Z.-M.; Zhang, Y.-Q.; Tu, Y.-Q.; Zhang, F.-M. J. Org. Chem. 2011, 76, 10173. (c) Nazef, N.; Davies, R. D. M.; Greaney, M. Org. Lett. 2012, 14, 3720.

(4) Selected recent examples and references cited therein: (a) Luo, J.; Jiang, Q.; Chen, H.; Tang, Q. RSC Adv. 2015, 5, 67901. (b) Parida, B. B.; Das, P. P.; Niocel, M.; Cha, J. K. Org. Lett. 2013, 15, 1780. (c) Yang, Y.; Ni, F.; Shu, W.-M.; Wu, A.-X. Tetrahedron 2014, 70, 6733. (d) Huang, S.; Kötzner, L.; De, K. C. J. Am. Chem. Soc. 2015, 137, 3446. (e) Mao, S.; Gao, Y.-R.; Zhang, S.-L.; Guo, D.-D.; Wang, Y.-Q. Eur. J. Org. Chem. 2015, 2015, 876. (f) Setzer, P.; Forcher, G.; Boeda, F.; Pearson-Long, M. S. M.; Bertus, P. Eur. J. Org. Chem. 2014, 171. (g) Mizar, P.; Wirth, T. Angew. Chem., Int. Ed. 2014, 53, 5993.

(5) For reviews, see: (a) de Alaniz, J.; Rovis, T. Synlett 2009, 2009, 1189. (b) Yetra, S. R.; Patra, A.; Biju, A. T. Synthesis 2015, 47, 1357. (c) Izquierdo, J.; Hutson, G. E.; Cohen, D. T.; Scheidt, K. A. Angew. Chem., Int. Ed. 2012, 51, 11686. For a book, see: (d) Gravel, M.; Holmes, J. M. In Comprehensive Organic Synthesis III; Knochel, P., Molander, G. A., Eds; Elsevier: Oxford, U.K., 2014; Vol. 4, pp 1384–1406.

(6) Wilde, M. M. D.; Gravel, M. Angew. Chem., Int. Ed. 2013, 52, 12651.

(7) (a) Galzerano, P.; Bencivenni, G.; Pesciaioli, F.; Mazzanti, A.; Giannichi, B.; Sambri, L.; Bartoli, G.; Melchiorre, P. *Chem. - Eur. J.* **2009**, *15*, 7846. (b) Sun, W.; Hong, L.; Liu, C.; Wang, R. *Tetrahedron: Asymmetry* **2010**, *21*, 2493. (c) Freund, M. H.; Tsogoeva, S. B. *Synlett* **2011**, *2011*, 503.

(8) (a) Ding, L.-Z.; Zhong, T.-S.; Wu, H.; Wang, Y.-M. *Eur. J. Org. Chem.* **2014**, 2014, 5139. (b) Sun, W.; Hong, L.; Zhu, G.; Wang, Z.; Wei, X.; Ni, J.; Wang, R. *Org. Lett.* **2014**, *16*, 544. (c) Sun, W.; Zhu, G.; Wu, C.; Hong, L.; Wang, R. *Chem. - Eur. J.* **2012**, *18*, 13959.

(9) (a) Lee, H. J.; Lim, J. W.; Yu, J.; Kim, J. N. *Tetrahedron Lett.* **2014**, 55, 1183. (b) Asahara, H.; Kida, T.; Hinoue, T.; Akashi, M. *Tetrahedron* **2013**, 69, 9428.

(10) (a) Zhao, S.; Lin, J.-B.; Zhao, Y.-Y.; Liang, Y.-M.; Xu, P.-F. Org. Lett. 2014, 16, 1802. (b) Wu, B.; Chen, J.; Li, M.-Q.; Zhang, J.-X.; Xu, X.-P.; Ji, S.-J.; Wang, X.-W. Eur. J. Org. Chem. 2012, 2012, 1318.
(c) Companyó, X.; Zea, A.; Alba, A.-N. R.; Mazzanti, A.; Moyano, A.; Rios, R. Chem. Commun. 2010, 46, 6953.

(11) (a) Scala, A.; Cordaro, M.; Grassi, G.; Piperno, A.; Barberi, G.; Cascio, A.; Risitano, F. *Bioorg. Med. Chem.* **2014**, *22*, 1063. (b) Robert, R.; Edwin, S. J. J. Chem. Res. Synopses **1990**, *2*, 48. (12) Compounds **3a** and **3b** are hard to separate on a silica gel column. Luckily, a single crystal of **3a** can be obtained by slowly evaporating the solvent from the mixture of **3a** and **3b** in ethyl acetate/ petroleum ether (see the Supporting Information).

(13) For reviews, see: (a) Shi, Z.; Zhang, C.; Tang, C.; Jiao, N. Chem. Soc. Rev. 2012, 41, 3381. (b) Wu, W.; Jiang, H. Acc. Chem. Res. 2012, 45, 1736. (c) Wendlandt, A. E.; Suess, A. M.; Stahl, S. S. Angew. Chem., Int. Ed. 2011, 50, 11062. For recent examples, see: (d) Liang, Y.-F.; Jiao, N. Angew. Chem., Int. Ed. 2014, 53, 548. (e) Miao, C.-B.; Zhang, M.; Tian, Z.-Y.; Xi, H.-T.; Sun, X.-Q.; Yang, H.-T. J. Org. Chem. 2011, 76, 9809. (f) Miao, C.-B.; Wang, Y.-H.; Xing, M.-L.; Lu, X.-W.; Sun, X.-Q.; Yang, H.-T. J. Org. Chem. 2013, 78, 11584.

(14) At present, we cannot obtain a single crystal to confirm the stereo configuration of 9.

(15) (a) Yang, Y.; Moinodeen, F.; Chin, W.; Ma, T.; Jia, Z.; Tan, C.-H. Org. Lett. **2012**, *14*, 4762. (b) Wang, G.-W.; Lu, Q.-Q.; Xia, J.-J. Eur. J. Org. Chem. **2011**, 2011, 4429.

(16) Zhang, P.; Bierer, D. E. J. Nat. Prod. 2000, 63, 643.

(17) Gao, G.-L.; Niu, Y.-N.; Yan, Z.-Y.; Wang, H.-L.; Wang, G.-W.; Shaukat, A.; Liang, Y.-M. J. Org. Chem. 2010, 75, 1305.

(18) (a) Sano, D.; Nagata, K.; Itoh, T. Org. Lett. 2008, 10, 1593.
(b) Lubin, H.; Tessier, A.; Chaume, G.; Pytkowicz, J.; Brigaud, T. Org. Lett. 2010, 12, 1496.

(19) (a) Aeberli, P.; Houlihan, W. J. J. Org. Chem. 1968, 33, 1640.
(b) Rajeswaran, W. G.; Labroo, R. B.; Cohen, L. A.; King, M. M. J. Org. Chem. 1999, 64, 1369.
(c) Takayama, H.; Shimizu, T.; Sada, H.; Harada, Y.; Kitajima, M.; Aimi, N. Tetrahedron 1999, 55, 6841.
(d) Nakagawa, M.; Kato, S.; Fukazawa, H.; Hasegawa, Y.; Miyazawa, J.; Hino, T. Tetrahedron Lett. 1985, 26, 5871.

(20) Kim, J. M.; Na, J. E.; Kim, J. N. *Tetrahedron Lett.* 2003, 44, 6317.
(21) Xu, X.-H.; Wang, X.; Liu, G.-K.; Tokunaga, E.; Shibata, N. Org. Lett. 2012, 14, 2544.

(22) (a) Nepali, K.; Singh, G.; Turan, A.; Agarwal, A.; Sapra, S.; Kumar, R.; Banerjee, U.; Verma, P. K.; Satti, N. K.; Gupta, M. K.; Suri, O. P.; Dhar, K. L. *Bioorg. Med. Chem.* **2011**, *19*, 1950. (b) Schäfer, I.; Opatz, T. *Synthesis* **2011**, *2011*, 1691.

(23) Phillips, E. M.; Riedrich, M.; Scheidt, K. A. J. Am. Chem. Soc. 2010, 132, 13179.